Monatshefte für Chemie Chemical Monthly Printed in Austria

Convenient and High-Yielding Preparations of Mono(5-carboxy-2-ethylpentyl) Phthalate and its Ring-Deuterated Isomer – The "Third" Major Metabolite of Bis(2-ethylhexyl) Phthalate

Hans-Detlev Gilsing^{1,*}, Jürgen Angerer², and Dietrich Prescher¹

¹ Institut für Dünnschichttechnologie und Mikrosensorik e.V., D-14513 Teltow, Germany

² Institut und Poliklinik für Arbeits-, Sozial- und Umweltmedizin der Universität Erlangen-Nürnberg, D-91054 Erlangen, Germany

Received September 7, 2004; accepted September 20, 2004 Published online January 18, 2005 © Springer-Verlag 2005

Summary. The synthesis of an oxidative major metabolite of bis(2-ethylhexyl) phthalate is described. The target molecule and its ring-deuterated isomer were obtained via acylation of the appropriate ω -hydroxy benzyl ester or the corresponding carboxylate with phthalic anhydride or phthalic anhydride-d₄. All transformation steps proceed with high yields.

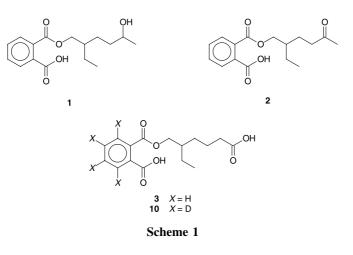
Keywords. Bis(2-ethylhexyl) phthalate; Oxidative metabolites; Carboxylic acids; Protecting groups; Ozonolysis.

Introduction

Due to their use as additives in many products of every day life phthalates can be found as ubiquitous contaminants in the environment [1]. Recently human exposure to phthalates has been proven unequivocally [2]. Several phthalates cause hepatic peroxisome proliferation [3] and testicular damage [4] in animal experiments. The relevance of these results with regard to human health is still debated controversially [5] so that further toxicological investigations must reveal potentially existing risks. The metabolism of bis(2-ethylhexyl) phthalate (*DEHP*) has been elucidated [6] and compounds 1, 2, and 3 have been identified as major metabolites (Scheme 1).

The major metabolites are formed by different ω - and (ω -1)-oxidation steps proceeding consecutively from mono(2-ethylhexyl) phthalate (*MEHP*) which is the

^{*} Corresponding author. E-mail: gilsing@idm-teltow.de



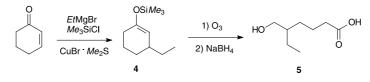
primary metabolite of *DEHP*. Compound **3** is metabolized further by degradation [7] with loss of a two-carbon fragment. The concentration of **3** detected in rats has been found to be time dependent and strongly related to the amount of *DEHP* or *MEHP* administered [8].

A new powerful method [9] of biological monitoring allows the measurement of exposure to *DEHP* by determination of its oxidative metabolites 1 and 2 in urine. This method needs the original compounds and their isotopically labelled analogues as reference materials. In the past it has been necessary to gain reference material from animal experiments until a synthetic access to 1 and 2 and their deuterated isomers has been developed [10]. We now report our efforts in the synthesis of 3 [11] and its deuterated isomer 10, which may also serve as biomarkers.

Results and Discussion

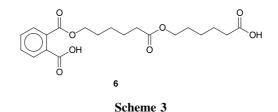
The retrosynthetic disconnection of the target molecule **3** leads to phthalic anhydride and **5** [12], which is a derivative of 6-hydroxycaproic acid. The introduction of the ethyl group was achieved by copper catalyzed conjugate addition [13] of the appropriate *Grignard* reagent to 2-cyclohexenone in the presence of chlorotrimethylsilane and ozonolytic cleavage [14] of the resulting silyl enol ether **4** [15]. These steps yielded **5** with very good overall yield (Scheme 2).

In order to investigate if direct acylation of 5 is possible some screening experiments with commercial 6-hydroxycaproic acid were done. Whereas the acylation of this compound with acetic anhydride is known [16] the reaction with phthalic anhydride showed only low conversion. Using pyridine as solvent gave 6 as single



Scheme 2

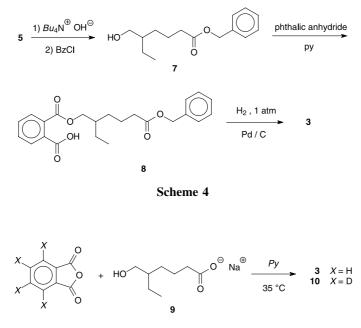
The "Third" Metabolite of Bis(2-ethylhexyl) Phthalate



product, which is formed by intermolecular esterification and was identified by comparison of its NMR spectra with published data [17] of similar structures (Scheme 3). Attempted acylation in chloroform with 4-(dimethylamino)pyridine as catalyst [18] was not successful.

As a conclusion of these results the benzyl group was choosen for the protection [19] of the carboxylic group of **5** because it can be removed easily in the presence of another ester group. The benzylation of **5** via the tetrabutylammonium salt [20] gave the hydroxy benzyl ester **7**. The target molecule **3** was obtained in high overall yield by acylation of **7** using an excess of phthalic anhydride and subjecting the resulting monophthalate **8** to hydrogenolysis [21] with Pd/C as catalyst (Scheme 4).

This reaction sequence provides an easy access to the target structure. In the view of the expensive deuterated anhydride, however, it is desired to avoid isotopically labelled intermediates. Therefore we checked the acylation [22] of ω -hydroxycarboxylate 9. Although it was necessary to increase the temperature slightly in order to dissolve the salt and the solution remained thick during the reaction acidic work-up gave 10 with 69% yield (Scheme 5).



Scheme 5

Experimental

IR spectra were recorded on a Perkin-Elmer 1430 spectrophotometer using the film technique. NMR spectra were recorded on a Varian Mercury plus-BB/4-nuc spectrometer operating at 400 MHz for ¹H and 100 MHz for ¹³C. High resolution mass spectra were recorded on a VG 7070 double focusing mass spectrometer using the CI and the NCI mode. Elemental analyses were carried out using a CE Instruments CHNS-Elemental Analyzer EA 1110. All new compounds gave satisfactory data of elemental analysis or high resolution mass spectrometry. All reactions except the neutralization of **5** were carried out under an atmosphere of Ar. 6-Hydroxycaproic acid and phthalic anhydride-d₄ (99.4%) were purchased from Aldrich and used as received. Purifications by column chromatography were performed on silica gel (Merck 63-200 mesh).

3-(Ethylcyclohex-1-enyloxy)trimethylsilane (4)

To a solution of ethylmagnesium bromide, prepared from 1.70 g of Mg turnings (70.0 mmol) and 5.23 cm³ of ethyl bromide (70.0 mmol) in 100 cm³ of anhydrous *THF* was added 0.51 g of CuBr · *Me*₂S (2.5 mmol) at -78° C. A solution of 20.88 cm³ of *HMPT* (120.0 mmol) in 100 cm³ of *THF* was then added dropwise. The mixture was stirred for 15 min, a solution of 3.85 cm³ of 2-cyclohexenone (39.8 mmol) and 12.68 cm³ of chlorotrimethylsilane (100.0 mmol) in 20 cm³ of *THF* was then added dropwise. The reaction mixture was stirred for 3 h at -78° C, and a solution of 14.05 cm³ of triethyl-amine (100.0 mmol) in 20 cm³ of *THF* was added. The mixture was stirred for additional 30 min, warmed up to room temperature, hydrolyzed with a saturated aqueous solution of NaHCO₃ and diluted with 100 cm³ of heptane. The filtrate was washed with $10 \times 20 \text{ cm}^3$ of a saturated aqueous solution of NaHCO₃ and dried (Na₂SO₄). The drying agent was filtered off, the solvent was evaporated under reduced pressure, and the product was isolated from the residue by Kugelrohr distillation. Yield 6.57 g (83%); colourless liquid; the analytical data agreed with the published ones [15].

5-(Hydroxymethyl)heptanoic acid (5)

A solution of 5.25 g of 4 (26.5 mmol) in 20 cm³ of anhydrous *Me*OH and 5 cm³ of CH₂Cl₂ was treated with O₃ at -78° C until a blue colour persisted. 1.00 g of sodium borohydride (26.5 mmol) was added in portions and the mixture was stirred for 1 h. Additional portions of 1.00 g of sodium borohydride (26.5 mmol) were then added, the mixture was warmed up to room temperature, and poured into 200 cm³ of a saturated aqueous solution of NaHCO₃. The organic layer was discarded and the aqueous layer was extracted with $3 \times 20 \text{ cm}^3$ of diethyl ether. The aqueous phase was then acidified with 400 cm³ of a 5% aqueous HCl, saturated with NaCl and extracted with $4 \times 100 \text{ cm}^3$ of CHCl₃. The combined extracts were dried (MgSO₄). The drying agent was filtered off and the solvent was evaporated under reduced pressure. Yield 4.04 g (96%); colourless oil; ¹H NMR (400 MHz, *DMSO*-d₆): $\delta = 0.81-0.86$ (m, CHCH₂CH₃), 1.10–1.65 (m, HOCH₂CH–(CH₂CH₃)CH₂CH₂CO₂H), 2.11–2.22 (m, CH₂CO₂H), 3.17–3.42 (m, CH₂OH), 4.25 (s, br, OH), 11.94 (s, br, CO₂H) ppm; ¹³C NMR (100 MHz, *DMSO*-d₆): $\delta = 10.9$, 22.0, 22.9, 29.7, 34.1, 41.3, 62.9, 174.4 ppm; IR (film): $\bar{\nu} = 3500-2400$, 2932, 2876, 1704, 1461, 1410, 1235, 1171, 1027, 755 cm⁻¹; HRMS (NH₃–CI) [M+H]⁺: calcd.: 161.1178, found: 161.1182.

Phthalic acid mono[5-(5-carboxy-pentyloxycarbonyl)-pentyl]ester (6, C₂₀H₂₆O₈)

A solution of 1.90 g of phthalic anhydride (12.8 mmol) and 0.95 g of 6-hydroxycaproic acid (7.2 mmol) in 100 cm³ of anhydrous pyridin was refluxed for 11 h, cooled, and stirred at room temperature for additional 40 h. The solvent was evaporated under reduced pressure and the residue was diluted with 50 cm^3 of CHCl₃, washed with $2 \times 20 \text{ cm}^3$ of a 1*M* aqueous HCl and $2 \times 20 \text{ cm}^3$ of H₂O and dried

The "Third" Metabolite of Bis(2-ethylhexyl) Phthalate

(MgSO₄). The drying agent was filtered off and the solvent was evaporated under reduced pressure. The product was isolated from the residue by column chromatography using ethylacetate:heptane = 1:1 as eluent. Yield 0.14 g (10%); pale yellow waxy solid; ¹H NMR (400 MHz, *DMSO*-d₆): δ = 1.23–1.68 (m, OCH₂CH₂CH₂CH₂CH₂CO₂CH₂CH₂CH₂CH₂CH₂CO₂H), 2.17–2.32 (m, 2 CH₂CO₂), 4.00–4.04 (m, OCH₂), 4.18–4.27 (m, OCH₂), 7.60–7.63 (m, 3H, Ar), 7.72–7.76 (m, 1H, Ar), 12.54 (s, br, 2 CO₂H) ppm; ¹³C NMR (100 MHz, *DMSO*-d₆): δ = 24.1, 24.9, 27.5, 27.6, 27.7, 27.8, 33.3, 33.5, 63.4, 64.8, 127.9, 128.5, 130.7, 131.0, 131.9, 132.2, 167.2, 167.7, 172.5, 174.0 ppm; IR (film): $\bar{\nu}$ = 3500–2500, 2940, 2868, 1721, 1699, 1600, 1579, 1284, 1259, 1125, 1072, 743 cm⁻¹.

5-(Hydroxymethyl)heptanoic acid benzyl ester (7, C₁₅H₂₂O₃)

A mixture of 1.64 cm³ of an 40% aqueous solution of tetrabutylammonium hydroxide (2.5 mmol) and 0.40 g of **5** (2.5 mmol) was stirred at 65°C until a clear solution appeared. The solvent was removed under reduced pressure and the residual oil was dried under high vacuum. 1.00 g of oil (2.5 mmol) was dissolved in 15 cm³ of anhydrous *DMF* and 0.32 cm³ of benzylchloride (2.8 mmol) were added and the mixture was stirred at room temperature for 24 h. The reaction mixture was poured into 150 cm³ of H₂O and was extracted with 3×50 cm³ of diethyl ether. The combined extracts were dried (Na₂SO₄). The drying agent was filtered off and the solvent was evaporated under reduced pressure. The product was isolated from the residue by column chromatography using ethylacetate:heptane = 2:3 as eluent. Yield 0.50 g (81%); colourless oil; ¹H NMR (400 MHz, *DMSO*-d₆): δ = 0.81 (t, *J* = 7.3 Hz, CHCH₂CH₃), 1.19–1.31 (m, HOCH₂CH(CH₂CH₃)CH₂), 1.54 (t, *J* = 7.3 Hz, CH₂CH₂CO₂CH₂Ar), 2.33 (t, *J* = 7.3 Hz, CH₂CO₂CH₂Ar), 3.26–3.30 (m, CH₂OH), 4.28 (t, *J* = 5.3 Hz, OH), 5.08 (s, OCH₂Ar), 7.31–7.37 (m, 5H, Ar) ppm; ¹³C NMR (100 MHz, *DMSO*-d₆): δ = 10.9, 21.9, 22.9, 29.5, 33.9, 41.2, 62.9, 65.2, 127.8, 127.9, 128.4, 136.3, 172.7 ppm; IR (film): $\bar{\nu}$ = 3450, 2959, 2931, 2875, 1733, 1456, 1158, 1029, 736, 696 cm⁻¹; HRMS (NH₃–NCI) [M – H]⁻: calcd.: 249.1491, found: 249.1495.

Phthalic acid mono(5-benzyloxycarbonyl-2-ethylpentyl)ester (8, C₂₃H₂₆O₆)

A solution of 1.07 g of phthalic anhydride (7.2 mmol) and 0.72 g of **7** (2.9 mmol) in 50 cm³ of anhydrous pyridine was stirred at room temperature for 72 h. The mixture was evaporated under reduced pressure and the residue was diluted with 100 cm³ of diethyl ether. The solution was washed with 3×20 cm³ of a 10% aqueous HCl, dried (Na₂SO₄), and evaporated. The product was isolated from the residue by column chromatography using ethylacetate:heptane = 5:1 as eluent. Yield 0.94 g (82%); colourless oil; ¹H NMR (400 MHz, *DMSO*-d₆): $\delta = 0.86$ (t, J = 7.3 Hz, CHCH₂CH₃), 1.24–1.39 (m, OCH₂CH(CH₂CH₃)CH₂CH₂CH₂CO₂CH₂Ar), 1.55–1.67 (m, OCH₂CH(CH₂CH₃)CH₂CH₂CH₂CH₂CO₂CH₂Ar), 4.09–4.16 (m, OCH₂CH), 5.08 (s, OCH₂Ar), 7.29–7.38 (m, 5H, Ar), 7.60–7.65 (m, 3H, Ar), 7.71–7.76 (m, 1H, Ar), 13.18 (s, br, CO₂H) ppm; 1³C NMR (100 MHz, *DMSO*-d₆): $\delta = 10.7$, 21.7, 23.1, 29.5, 33.6, 37.8, 65.2, 67.0, 127.9, 128.1, 128.3, 128.7, 130.9, 131.1, 132.3, 136.2, 167.5, 168.0, 172.6 ppm; IR (film): $\bar{\nu} = 3400-2500$, 3034, 2961, 2877, 1725, 1698, 1600, 1580, 1456, 1383, 1285, 1259, 1123, 1072, 742, 697, 639 cm⁻¹; HRMS (NH₃–NCI) [M]⁻⁻: calcd.: 398.1729, found: 398.1743.

Phthalic acid mono(5-carboxy-2-ethylpentyl)ester (3, C₁₆H₂₀O₆)

To a solution of 0.30 g of **8** (0.8 mmol) in 25 cm³ of anhydrous *Me*OH was added 0.03 g of Pd/C (10% Pd). Hydrogen was passed through at room temperature for 8 h and the reaction mixture was stirred under 1 atm of hydrogen for additional 16 h. The mixture was filtered and the filtrate was evaporated under reduced pressure. The residue was diluted with 100 cm³ of diethyl ether and the solution was washed with 2×100 cm³ of a saturated aqueous solution of NaHCO₃. The organic layer was discarded

and the aqueous layer was acidified, saturated with NaCl, and extracted with $3 \times 100 \text{ cm}^3$ of CHCl₃. The combined extracts were dried (Na₂SO₄) and evaporated to give the pure product. Yield 0.18 g (78%); colourless oil; ¹H NMR (400 MHz, *DMSO*-d₆): $\delta = 0.88$ (t, J = 7.3 Hz, CHCH₂CH₃), 1.23–1.40 (m, OCH₂CH(CH₂CH₃)CH₂CH₂CH₂CO₂H), 1.41–1.58 (m, OCH₂CH(CH₂CH₃)CH₂CH₂CO₂H), 1.60–1.69 (m, OCH₂CH(CH₂CH₃)), 2.15–2.23 (m, CH₂CO₂H), 4.09–4.17 (m, OCH₂CH), 7.60–7.66 (m, 3H, Ar), 7.72–7.76 (m, 1H, Ar), 12.56 (s, br, 2 CO₂H) ppm; ¹³C NMR (100 MHz, *DMSO*-d₆): $\delta = 10.7$, 21.7, 23.1, 29.6, 33.8, 37.9, 67.0, 128.1, 128.7, 130.9, 131.1, 132.2, 132.3, 167.5, 168.0, 174.3 ppm; IR (film): $\bar{\nu} = 3400-2500$, 2961, 2877, 2665, 1721, 1698, 1600, 1580, 1412, 1383, 1284, 1259, 1124, 1073, 742, 705, 675, 640 cm⁻¹; HRMS (NH₃–NCI) [M – H]⁻: calcd.: 307.1182, found: 307.1168.

Sodium 5-(hydroxymethyl)heptanoate (9, C₈H₁₅NaO₃)

To a solution of 0.13 g of NaOH (3.1 mmol) in 15 cm³ of distilled H₂O was added 0.50 g of **5** (3.1 mmol) and the mixture was stirred at 65°C until a homogenous phase appeared. The solvent was evaporated and the residue solidified upon drying under high vacuum. Yield 0.57 g (100%); colourless solid; mp 93–94°C; IR (film): $\bar{\nu} = 3170, 2959, 2935, 2874, 1571, 1459, 1379, 1062, 883, 739 cm⁻¹.$

Preparation of 3 via Acylation of 9

To a solution of 0.46 g of phthalic anhydride (3.1 mmol) in 25 cm³ of anhydrous pyridine was added 0.57 g of **9** (3.1 mmol), the mixture was stirred at 35°C for 1 h to give a thick solution which was stirred at room temperature for additional 72 h. The solution was poured into 250 cm³ of a 10% aqueous HCl. The aqueous solution was saturated with NaCl and extracted with 3×100 cm³ of CHCl₃. The combined extracts were dried (Na₂SO₄). The drying agent was filtered off and the solvent was evaporated under reduced pressure. The residue was partitionated between 200 cm³ of a saturated aqueous solution of NaHCO₃ and 100 cm³ of diethyl ether and the organic layer was discarded. The aqueous layer was acidified, saturated with NaCl, and extracted with 3×100 cm³ of CHCl₃. The combined extracts were dried to give the pure product. Yield 0.63 g (66%).

Phthalic acid (3,4,5,6-²H₄) mono(5-carboxy-2-ethylpentyl)ester (10, C₁₆H₁₆D₄O₆)

To a solution of 0.42 g of phthalic anhydride-d₄ (2.7 mmol) in 35 cm³ of anhydrous pyridine was added 0.50 g of **9** (2.7 mmol), the mixture was stirred at 35°C for 1 h to give a thick solution which was stirred at room temperature for additional 72 h. After work-up as described above the pure product was obtained. Yield 0.59 g (69%); colorless oil; ¹H NMR (400 MHz, *DMSO*-d₆): δ = 0.88 (t, *J* = 7.3 Hz, CHCH₂CH₃), 1.24–1.42 (m, OCH₂CH(CH₂CH₃)CH₂CH₂CH₂-CO₂H), 1.50–1.58 (m, OCH₂CH (CH₂CH₃)CH₂CH₂CH₂CH₂CO₂H), 1.60–1.68 (m, OCH₂CH(CH₂CH₃)), 2.19–2.23 (m, CH₂CO₂H), 4.11–4.13 (m, OCH₂CH), 12.61 (s, br, 2 CO₂H) ppm; ¹³C NMR (100 MHz, *DMSO*-d₆): δ = 10.7, 21.7, 23.1, 29.6, 33.9, 37.9, 67.0, 132.2, 132.3, 167.6, 168.0, 174.4 ppm; IR (film): $\bar{\nu}$ = 3400–2500, 2962, 2936, 1697, 1557, 1432, 1235, 1077, 941, 751, 666 cm⁻¹; HRMS (NH₃–NCI) [M – H]⁻: calcd.: 311.1433, found: 311.1436.

Acknowledgements

The authors thank Dr. K. Seiler, ASCA Berlin, for recording the NMR spectra, Dr. M. Bartoszek, ACA Berlin, for providing the HRMS data, Dr. U. Gross, Humboldt University Berlin, for performing the elemental analyses, Dr. F. Theil, ASCA Berlin, for his help during the ozonolysis, and E. Tews and D. Stolle for conducting the preparative work. Financial support by the Deutsche Forschungsgemeinschaft under AN107/16-1 is gratefully acknowledged. H.-D. Gilsing dedicates this article to the memory of his beloved mother.

References

- a) Muszkat L, Bir L, Raucher D (1997) Bull Environ Contam Toxicol 58: 348; b) Tsumura Y, Ishimitsu S, Saito I, Sakai H, Kobayashi Y, Tonogai Y (2001) Food Addit Contam 18: 449; c) Suzuki T, Yaguchi K, Suzuki S, Suga T (2001) Environ Sci Technol 35: 3757
- [2] a) Koch HM, Rossbach B, Drexler H, Angerer J (2003) J Environ Res 93: 177; b) Koch HM, Drexler H, Angerer J (2003) Int J Hyg Environ Health 206: 77; c) Koch HM, Drexler H, Angerer J (2003) Umweltmed Forsch Prax 8: 15; d) Barr DB *et al.* (2003) Environ Health Perspect 111: 1148
- [3] Elcombe CR, Mitchell AM (1986) Environ Health Perspect 70: 211
- [4] Gray TJB, Gangolli SD (1986) Environ Health Perspect 65: 229
- [5] a) David RM (2000) Environ Health Perspect 108: A440; b) Kohn MC, Parham F, Masten SA, Portier CJ, Shelby MD, Brock JW, Needham LL (2000) Environ Health Perspect 108: A440; c) Melnick RL (2001) Environ Health Perspect 109: 437; d) Lovekamp-Swan T, Davis BJ (2003) Environ Health Perspect 111: 139
- [6] a) Albro PW, Thomas R, Fishbein L (1973) J Chromatogr 76: 321; b) Daniel JW, Bratt H (1974) Toxicology 2: 51; c) Chu I, Villeneuve DC, Secours V, Franklin C, Rock G, Viau A (1978) Drug Metab Dispos 6: 146; d) Albro PW, Tondeur I, Marbury D, Jordan S, Schroeder J, Corbett JT (1983) Biochim Biophys Acta 760: 283; e) Schmid P, Schlatter C (1985) Xenobiotica 15: 251; f) Albro PW, Lavenhar SR (1989) Drug Metab Rev 21: 13; g) Koch HM, Bolt HM, Angerer J (2004) Arch Toxicol 78: 123
- [7] a) Albro PW, Corbett JT, Schroeder J, Reddy JK (1987) Biochim Biophys Acta 923: 196;
 b) Dirven HAAM, van den Broek PHH, Peeters MCE, Peters JGP, Mennes WC, Blaauboer BJ, Noordhoek J, Jongeneelen FJ (1993) Biochem Pharmacol 45: 2425
- [8] Lhuguenot JC et al. (1985) Toxicol Appl Pharmacol 80: 11
- [9] Koch HM, Gonzalez-Reche LM, Angerer J (2003) J Chromatogr B 784: 169
- [10] a) Gilsing H-D, Angerer J, Prescher D (2002) Monatsh Chem 133: 1147; b) Gilsing H-D, Angerer J, Prescher D (2003) Monatsh Chem 134: 1207
- [11] Sjöberg P, Bondesson U, Gray TJB, Plöen L (1986) Acta Pharmacol Toxicol 58: 225
- [12] a) Kaulen J, Schäfer H-J (1982) Tetrahedron 38: 3299; b) Gomez-Bengoa E, Heron NM, Didiuk MT, Luchaco CA, Hoveyda AM (1998) J Am Chem Soc 120: 7649
- [13] a) Horiguchi Y, Matsuzawa S, Nakamura E, Kuwajima I (1986) Tetrahedron Lett 27: 4025;
 b) Nakamura E (1994) Me₃SiCl Accelerated Conjugate Additions. In: Taylor RJK (ed) Organocopper Reagents A Practical Approach. Oxford Press, New York, p 129
- [14] Clark RD, Heathcock CH (1976) J Org Chem 41: 1396
- [15] Denmark SE, Dappen MS, Sear NL, Jacobs RT (1990) J Am Chem Soc 112: 3466
- [16] Kanemitsu T, Wong C-H, Kanie O (2002) J Am Chem Soc 124: 3591
- [17] MacDonald RT, Pulapura SK, Svirkin YY, Gross RA, Kaplan DL, Akkara J, Swift G, Wolk S (1995) Macromolecules 28: 73
- [18] Höfle G, Steglich W, Vorbrüggen H (1978) Angew Chem 90: 602
- [19] Kocienski PJ (2004) Protecting Groups. Thieme, Stuttgart, p 409
- [20] Hum G, Wooler K, Lee J, Taylor SD (2000) Can J Chem 78: 642
- [21] Griesbeck AG, Nerowski F, Lex J (1999) J Org Chem 64: 5213
- [22] Goto G, Okamoto K, Okutani T, Imada I (1985) Chem Pharm Bull 33: 4422

Verleger: Springer-Verlag GmbH, Sachsenplatz 4–6, 1201 Wien, Austria. – Herausgeber: Österreichische Akademie der Wissenschaften, Dr.-Ignaz-Seipel-Platz 2, 1010 Wien, und Gesellschaft Österreichischer Chemiker, Eschenbachgasse 9, 1010 Wien, Austria. – Redaktion: Getreidemarkt 9/163-OC, 1060 Wien, Austria. – Satz und Umbruch: Thomson Press Ltd., Chennai, India. – Offsetdruck: Manz Crossmedia, 1051 Wien, Austria. – Verlagsort: Wien. – Herstellungsort: Wien. – Printed in Austria.